Adeno-Associated Virus Antibody Profiles in Newborns, Children, and Adolescents[∇]

Roberto Calcedo, Hiroki Morizono, Lili Wang, Robert McCarter, Jianping He, David Jones, Mark L. Batshaw, and James M. Wilson **

Gene Therapy Program, Department of Pathology and Laboratory Medicine, University of Pennsylvania, Philadelphia, Pennsylvania 19104¹;

Center for Genetic Medicine Research, Children's Research Institute, Children's National Medical Center,

Washington, DC 20010²; and Center for Clinical and Community Research, Children's Research Institute,

Children's National Medical Center, Washington, DC 20010³

Received 21 April 2011/Returned for modification 6 June 2011/Accepted 8 July 2011

Neutralizing antibodies (NAb) to an adeno-associated virus (AAV) vector due to previous natural infection with wild-type AAV can significantly limit gene transfer. NAb titers to AAV serotype 2 (AAV2) and AAV8 in human subjects (0 to 18 years) were studied. NAb prevalence is moderate at birth, decreases markedly from 7 to 11 months, and then progressively increases through childhood and adolescence.

Adeno-associated viruses (AAVs) are replication-defective members of the family *Parvoviridae* that have been widely used as vectors for gene therapy. Vectors based on AAV serotype 2 (AAV2) have been evaluated in both animal models and humans. The discovery of new serotypes, including AAV8 (6), has provided alternative vectors with substantial advantages over AAV2 in terms of gene transfer efficiency (4, 11, 13, 18), lower prevalence of neutralizing antibodies (NAbs) in the human population (1, 3), and less propensity to activate T cells for capsid proteins (14, 17).

It has been shown that preexisting NAbs to the viral vector limits effective gene transfer in a way that is influenced by the route of administration and organ targeted. Several studies have shown that even low levels of AAV NAbs can reduce gene transfer into the liver following intravascular delivery (7, 16) in the context of potential treatments of several genetic disorders, including hemophilia B and ornithine transcarbamy-lase deficiency.

The goal of this study was to evaluate the prevalence of NAbs to AAV2 and AAV8 in plasma from newborns, children, and adolescents to determine the ideal age interval for gene therapy intervention, which would be when the prevalence of AAV NAb is the lowest.

Plasma samples from 752 anonymous human subjects of different age groups (Table 1) were obtained from the Division of Laboratory Medicine at Children's National Medical Center (Washington, DC). Samples were heat inactivated at 56°C for 30 min and analyzed for Nab to AAV2 or AAV8 by an in vitro transduction inhibition assay (3). NAb titers were determined for each sample, and data were recorded as counts of positive responses among totals evaluated by vector, age, and dilution and used to estimate the prevalence of vector transduction inhibition at plasma dilutions of ≥ 1.5 , ≥ 1.10 , ≥ 1.20 , and ≥1:40 (Fig. 1). The NAb titer was reported as the highest plasma dilution that inhibited AAV transduction of Huh7 cells by 50% or more compared with that for the naive serum control. The limit of detection of the assay was 1:5. Stratified contingency table analyses and negative binomial regression models in the Stata 11 software program (12), appropriate for count-type data, were used to evaluate the impact of age and AAV serotype on the prevalence of seropositivity based on an AAV NAb titer equal to or greater than 1:20 (Table 1 and Table 2).

Based on the raw data shown in Fig. 1, NAbs with a plasma dilution of ≥1:5 were present at birth in 59% of subjects for AAV2 and in 36% for AAV8. Nineteen percent of neonates

TABLE 1. Average prevalence of NAb (titer of ≥1:20) by age in anonymous serum samples from Children's National Medical Center

Group	Age (yr)	No. of samples:		% prevalence	Relative	95% confidence	P value
		Tested	Positive	% prevalence	prevalence	interval	1 value
Infants ^a	<1	175	31	15			
Toddlers	1-<3	83	13	13.5	0.9	0.49, 1.64	0.72
Children Adolescents	3–18	350	96	21.5	1.43	0.99, 2.07	0.052

^a Reference group for comparisons of relative prevalence.

^{*} Corresponding author. Mailing address: Gene Therapy Program, Department of Pathology and Laboratory Medicine, University of Pennsylvania, Philadelphia, PA 19104. Phone: (215) 898-0226. Fax: (215) 494-5444. E-mail: wilsonjm@mail.med.upenn.edu.

[▽] Published ahead of print on 20 July 2011.

Vol. 18, 2011 NOTES 1587

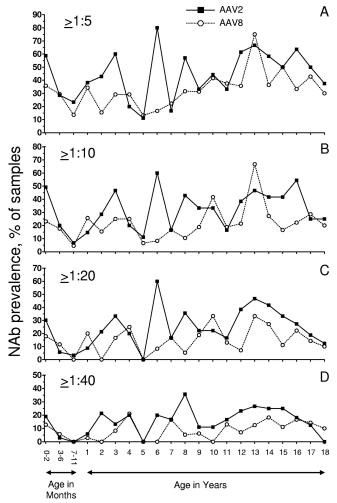


FIG. 1. Distribution of the prevalence of neutralizing antibodies (NAbs) against adeno-associated virus (AAV) types 2 and 8 in 751 (AAV2, n=353; AAV8, n=398) plasma samples from subjects with ages ranging from 1 day to 18 years. Samples were considered positive if serum dilutions of ≥ 1.5 (A), ≥ 1.10 (B), ≥ 1.20 (C), or ≥ 1.40 (D) inhibited vector transduction by $\geq 50\%$.

had plasma dilution titers of ≥1:40 for AAV2 and 13% for AAV8. Prevalence of NAbs to both AAV serotypes declined significantly after birth, reaching a nadir in the 7- to 11-month group, probably due to a drop in maternal antibody levels. These results are consistent with our statistical model based on the negative binomial that indicated a rapid decline in NAb prevalence during the first 6 months of life followed by a gradual increase with age thereafter, using a NAb titer of 1:20

TABLE 2. Average prevalence of NAb (titer of ≥ 1:20) by AAV serotype in anonymous serum samples from Children's National Medical Center

AAV	No. of samples:		%	Relative	95% confidence	P value
serotype	Tested	Positive	prevalence	prevalence	interval	1 value
2 ^a	275	78	22.1			
8	333	62	15.7	0.71	0.53, 0.96	0.025

^a Reference group for comparisons of relative prevalence.

in these and subsequent statistical analyses. Table 1 shows the average prevalence was 15% in infants (ages < 1 year), 14% in toddlers (ages 1 to <3 years), and 21% in older children (ages 3 to 18 years), the latter increase from infancy nearly reaching statistical significance (P = 0.052). Table 2 compares the average NAb prevalence across all ages for AAV2 (22%) and AAV8 (15%) and indicates that this difference achieves statistical significance (P = 0.025). Previous studies indicated the potential intrauterine transmission of maternal AAV into the fetus due the high susceptibility to infection of the trophoblast by AAV (2, 10) and the possible transmission of AAV during vaginal delivery (5, 15). Although our serological analysis does not indicate a persistent humoral immune response to AAV after birth, as would be expected if the newborns were infected at birth, it indicates an AAV infection after 1 year of age, with a peak at 3 years of age. This serologic pattern closely follows that of the adenovirus as described previously (9) and is consistent with the acquisition of AAV as a consequence of adenovirus infection.

Recent studies in monkeys have shown that very low levels of preexisting NAb to AAV8 can abrogate AAV8-mediated liver transduction (8, 16). In studies of liver-directed gene therapy, we have shown that an AAV8 NAb titer of 1:20 is sufficient to reduce transduction considerably and to redirect vector DNA to the spleen (17a). Our data suggest that 70% and 82% of newborns have titers below 1:20 (Fig. 1C) and would be suitable subjects for systemic delivery of AAV2 or AAV8, respectively. This percentage would increase to 97% for AAV2 and 100% for AAV8 if genetic intervention is delayed to 7 to 11 months of age.

In summary, our data indicate that the best age for an early gene therapy intervention with an AAV vector would be between 7 and 11 months of age and that after 3 years of age AAV8 would be a better delivery vector than AAV2 due to its lower NAb prevalence. AAV-mediated gene therapy on patients of any age clearly will require careful screening for pre-existing AAV NAbs due to the wide range of seroprevalences observed in the study. Our data also suggest that natural infections with AAV occur soon after the infant loses humoral protection due to passive transfer and remain stable until adolescence, when there is an apparent increase in infections.

This work was funded by grants to J.M.W. (NICHD P01 HD 057247-03) and to M.L.B. (NICHD U54061221) and by the Kettering Family Foundation. This research was also supported by awards UL1RR031988 and P30HD40677 from the NIH National Center for Research Resources and NIH Intellectual and Developmental Disabilities Research Center, respectively.

The contents of this work are solely the responsibility of the authors and do not necessarily represent the official views of the National Institutes of Health.

We thank the Immunology Core personnel (University of Pennsylvania, Gene Therapy Program), Qiuyue Qin and Surina Boyd, for their help analyzing plasma samples by the neutralizing antibody assay and the staff of Laboratory Medicine at Children's National Medical Center for obtaining specimens.

R.C., H.M., L.W., R.M., and M.L.B. have no conflicts of interest to declare. J.M.W. is a consultant to ReGenX Holdings and is a founder of, holds equity in, and receives a grant from affiliates of ReGenX Holdings; in addition, he is an inventor on patents licensed to various biopharmaceutical companies, including affiliates of ReGenX Holdings.

1588 NOTES CLIN. VACCINE IMMUNOL.

REFERENCES

- Boutin, S., et al. 2010. Prevalence of serum IgG and neutralizing factors against adeno-associated virus (AAV) types 1, 2, 5, 6, 8, and 9 in the healthy population: implications for gene therapy using AAV vectors. Hum. Gene Ther. 21:704–712.
- Burguete, T., et al. 1999. Evidence for infection of the human embryo with adeno-associated virus in pregnancy. Hum. Reprod. 14:2396–2401.
- Calcedo, R., L. H. Vandenberghe, G. Gao, J. Lin, and J. M. Wilson. 2009. Worldwide epidemiology of neutralizing antibodies to adeno-associated viruses. J. Infect. Dis. 199:381–390.
- Davidoff, A. M., et al. 2005. Comparison of the ability of adeno-associated viral vectors pseudotyped with serotype 2, 5, and 8 capsid proteins to mediate efficient transduction of the liver in murine and nonhuman primate models. Mol. Ther. 11:875–888.
- Friedman-Einat, M., et al. 1997. Detection of adeno-associated virus type 2 sequences in the human genital tract. J. Clin. Microbiol. 35:71–78.
- Gao, G. P., et al. 2002. Novel adeno-associated viruses from rhesus monkeys as vectors for human gene therapy. Proc. Natl. Acad. Sci. U. S. A. 99:11854– 11859
- Hurlbut, G. D., et al. 2010. Preexisting immunity and low expression in primates highlight translational challenges for liver-directed AAV8-mediated gene therapy. Mol. Ther. 18:1983–1994.
- Jiang, H., et al. 2006. Effects of transient immunosuppression on adenoassociated, virus-mediated, liver-directed gene transfer in rhesus macaques and implications for human gene therapy. Blood 108:3321–3328.
- 9. Parks, W. P., D. W. Boucher, J. L. Melnick, L. H. Taber, and M. D. Yow.

- 1970. Seroepidemiological and ecological studies of the adenovirus-associated satellite viruses. Infect. Immun. 2:716–722.
- 10. Parry, S., et al. 1998. Transduction of human trophoblastic cells by replication-deficient recombinant viral vectors. Promoting cellular differentiation affects virus entry. Am. J. Pathol. 152:1521–1529.
 11. Sarkar, R., et al. 2004. Total correction of hemophilia A mice with canine
- Sarkar, R., et al. 2004. Total correction of hemophilia A mice with canine FVIII using an AAV 8 serotype. Blood 103:1253–1260.
- StataCorp. 2009. Stata statistical software: release 11, 11th ed. StataCorp LP, College Station, TX.
- Sun, B., et al. 2005. Efficacy of an adeno-associated virus 8-pseudotyped vector in glycogen storage disease type II. Mol. Ther. 11:57–65.
- Vandenberghe, L. H., et al. 2006. Heparin binding directs activation of T cells against adeno-associated virus serotype 2 capsid. Nat. Med. 12:967–971.
- Walz, C. M., et al. 1998. Detection of infectious adeno-associated virus particles in human cervical biopsies. Virology 247:97–105.
- Wang, L., et al. 2010. The pleiotropic effects of natural AAV infections on liver-directed gene transfer in macaques. Mol. Ther. 18:126–134.
- Wang, L., J. Figueredo, R. Calcedo, J. Lin, and J. M. Wilson. 2007. Crosspresentation of adeno-associated virus serotype 2 capsids activates cytotoxic T cells but does not render hepatocytes effective cytolytic targets. Hum. Gene Ther. 18:185–194.
- 17a. Wang, L., et al. 8 June 2011. Impact of pre-existing immunity on gene transfer to nonhuman primate liver with adeno-associated virus 8 vectors. Hum. Gene Ther. [Epub ahead of print.] doi:10.1089/hum.2011.031.
- Ziegler, R. J., et al. 2007. Correction of the biochemical and functional deficits in fabry mice following AAV8-mediated hepatic expression of alphagalactosidase A. Mol. Ther. 15:492–500.